



MOMORDICA CHARANTIA PROTECTS AGAINST CARDIAC DAMAGE IN STREPTOZOTOCIN-INDUCED DIABETIC WISTAR RATS

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ABSTRACT

Diabetes mellitus is one of the most important world health problems, especially in developing countries where prevalence and incidence rates are highest. Diabetic patients are particularly prone to cardiovascular diseases including hypertension, atherosclerosis, diabetic cardiomyopathy, congestive heart failure and cardiac autonomic neuropathy. The present study investigated the effects of *Momordica charantia* (*M. charantia*) on histological changes of the left ventricle of the heart in streptozotocin-induced diabetic Wistar rats. Forty healthy adult Wistar rats of both sexes were randomly assigned into five groups A, B, C, D and E of eight rats each. Group A were the control (normal rats); B were the experimentally-induced diabetic rats; C were diabetic rats treated with methanolic extracts of *M. charantia* for two weeks (withdrawal group); D were diabetic rats treated with methanolic extracts of *M. charantia* for four weeks. E was diabetic rats treated with glimepiride for four weeks. Tissues were harvested, processed routinely in paraffin wax and stained with routine and special stains. Histological studies revealed disorganization of myofibril in the left ventricle of diabetic rats. Histochemical analysis also revealed abnormal deposition of glycogen in left ventricle of diabetic rats. *M. charantia* and glimepiride attenuated the morphological alterations and reduced the glycogen deposits.

Keywords: Diabetes, *Momordica charantia*, Histology, Heart, Left ventricle

INTRODUCTION

Diabetes mellitus is a serious metabolic disorder with micro and macro-vascular complications that result in a significant morbidity and mortality. Increasing proportion of the aging population, consumption of calorie rich diet, obesity and sedentary lifestyle have led to a tremendous increase in the number of diabetics worldwide¹. Diabetes mellitus is one of the most important world health problems, especially in developing countries where prevalence and incidence rates are highest. Diabetic patients are particularly prone to cardiovascular diseases including hypertension, atherosclerosis, diabetic cardiomyopathy, congestive heart failure and cardiac autonomic neuropathy². Coronary atherosclerosis and cardiomyopathy occur as a result of the metabolic abnormalities associated with diabetes³. These physical changes require years to develop in human following the onset of chronic hyperglycemia⁴. Bradycardia has also been noted to be a consistent feature when evaluating diabetic rat models^{5,6,7}. Heart rate variability in diabetes is commonly attributed to associated neuropathy⁸. Carnitine deficiency is often encountered in Diabetes mellitus^{9,10}. Structural changes in myocardial mitochondria have been associated with diabetes and concomitant carnitine deficiency⁵.

Momordica charantia (Linn Family: Cucurbitaceae) is one of the popular herbs that grow in different regions of Nigeria. It is commonly called Bitter melon, Bittergourd, Balsam pear. Bittergourd is known in some tribes of Nigeria as Ejirin wewe (Yoruba) Okban, Ndeme (Igbo) and Garafun (Hausa). It's a slender, climbing annual vine with long-stalked leaves and yellow, solitary male and female flowers borne in the leaf axils. The fruit looks like a warty gourd, usually oblong and resembling a small cucumber. The young fruit is emerald green, turning to orange-yellow when ripe. At maturity, the fruit splits into three irregular valves that curl backwards and release numerous reddish-brown or white seeds encased in scarlet arils. The Latin name *Momordica* means "to bite,"

referring to the jagged edges of the leaves, which appear as if they have been bitten. All parts of the plant, including the fruit, taste very bitter¹¹.

Various parts of *M. charantia* such as the seed, fruit and even the whole plant has been reported to have beneficial effects in prevention and treatment of many diseases in folkloric medicine, especially in the treatment of DM in individuals with non-insulin dependent diabetes^{12,13}. It has hypoglycaemic properties as it significantly suppressed the rise in blood glucose concentrations in albino rats^{12,14}. The first clinical study into the influence of the fresh juice of bittergourd on the management of DM was by Akhtar¹⁵. These findings suggested the intervention would effectively treat all symptoms of diabetes including polyuria, polydipsia, and polyphagia. Sarkar *et al.*,¹⁶ and Miura *et al.*,¹⁷ indicated that the fresh bitter-gourd juice caused a significant reduction in plasma glucose concentration, and an improvement in the response to an oral glucose load. Bitter melon contains an array of biologically active plant chemicals including triterpenes, proteins and steroids. In addition, a protein found in bitter melon, momordin, has clinically demonstrated anticancerous activity against Hodgkin's lymphoma in animals. Other proteins in the plant, alpha- and beta-momorcharin and cucurbitacinB, have been tested for possible anticancerous effects¹⁸. In numerous studies, at least three different groups of constituents found in all parts of bitter melon have clinically demonstrated hypoglycemic (blood sugar lowering) properties or other actions of potential benefit against diabetes mellitus¹⁹. These chemicals that lower blood sugar include a mixture of steroidal saponins known as charantins, insulin-like peptides, and alkaloids. The hypoglycemic effect is more pronounced in the fruit of bitter melon where these chemicals are found in greater abundance. The present study examined the effects of *M. charantia* on histological changes of the left ventricle of the heart in streptozotocin-induced diabetic Wistar rats and compared the

effects with those of glimepiride, an oral blood-glucose-lowering drug of the sulfonylurea class.

MATERIALS AND METHODS

Animal care

Forty healthy adult Wistar rats of both sexes, with average weight of 134.4g were used for the experiment. The rats were bred in the animal holding of College of Health Sciences, Obafemi Awolowo University, Ile-Ife. They were maintained on standard rat pellet (Cafsed, Ibadan, Nigeria) and water was provided *ad libitum*.

The animals were randomly assigned into five groups A, B, C, D and E of eight rats each. Group A were the control normal rats administered intraperitoneally with 0.1M sodium citrate buffer, group B were the rats induced with single dose of 65mg/kg streptozotocin and administered with 10% tween 80 for four weeks after four weeks of diabetic stabilization, group C and D were the experimentally-induced diabetic rats treated with methanolic extracts of *Momordica charantia* (100 mg/kg/day) dissolved in 10% tween 80 for two weeks (withdrawal group) and four weeks respectively after four weeks of diabetic stabilization, group E were the diabetic rats treated with a standard diabetic drug (2 mg/kg/day of glimepiride) dissolved in 10% tween 80 for four weeks after four weeks of diabetic stabilization.

The animals received humane treatment as outlined in the "Care and Management of Laboratory Animals" published by the National Institute of Health²⁰ with ethical clearance number (IRB/IEC 00005422)

Plant material:

Matured leaves of *Momordica charantia* (Cucurbitaceae) were collected during the raining season from suburban villages of Ile-Ife metropolis in Osun State of Nigeria. The leaves were taken to the Herbarium in the Department of Botany, Obafemi Awolowo University, Ile-Ife to confirm identification and a voucher specimen number (UHI 16510) was placed in the Herbarium.

Preparation of methanolic extract of *M. charantia*:

Leaves of *Momordica charantia* were air dried and powdered in a warring blender. A 765 g of the powdered leaves were extracted in 1.95 L of absolute methanol for 72 hours with intermittent shaking and filtered. The filtrate were concentrated *in vacuo* at 35°C using a vacuum rotary evaporator (Büchi Rotavapor R110, Schweiz). The extract were partitioned between water and dichloromethane, the dichloromethane fraction (5.94%) was oven-dried at 37°C and stored until it is ready to be used. Aliquot portions of the extract were weighed and dissolved in 10% tween 80 for use on each day of the experiment.

Induction of diabetes:

Diabetes mellitus was experimentally-induced in groups B, C, D and E by a single intraperitoneal injection of 65 mg/kg body weight of streptozotocin (Tocris Bioscience, UK) dissolved in 0.1M sodium citrate buffer (pH 6.3). Diabetes was confirmed in animals 48 hours after induction, by

determining fasting blood glucose level using a digital glucometer (Accu-chek® Advantage, Roche Diagnostic, Germany) consisting of a digital meter and the test strips using blood samples obtained from the tail vein of the rats. Animals in group A were given equal volume of citrate buffer used in dissolving streptozotocin intraperitoneally.

Administration of extract and anti-diabetic drug:

Methanolic extracts of the leaves of *M. charantia* (100 mg/kg) was dissolved in 10% tween 80 and administered daily (orally) by gastric intubation to the rats in groups C and D for 2 and 4 weeks respectively. The standard antidiabetic drug (glimepiride 2 mg/kg) was administered daily to group E rats for four weeks²¹, while the rats in group B were administered with the vehicle used in dissolving the extract and glimepiride (10% tween 80).

Sacrifice of the animals:

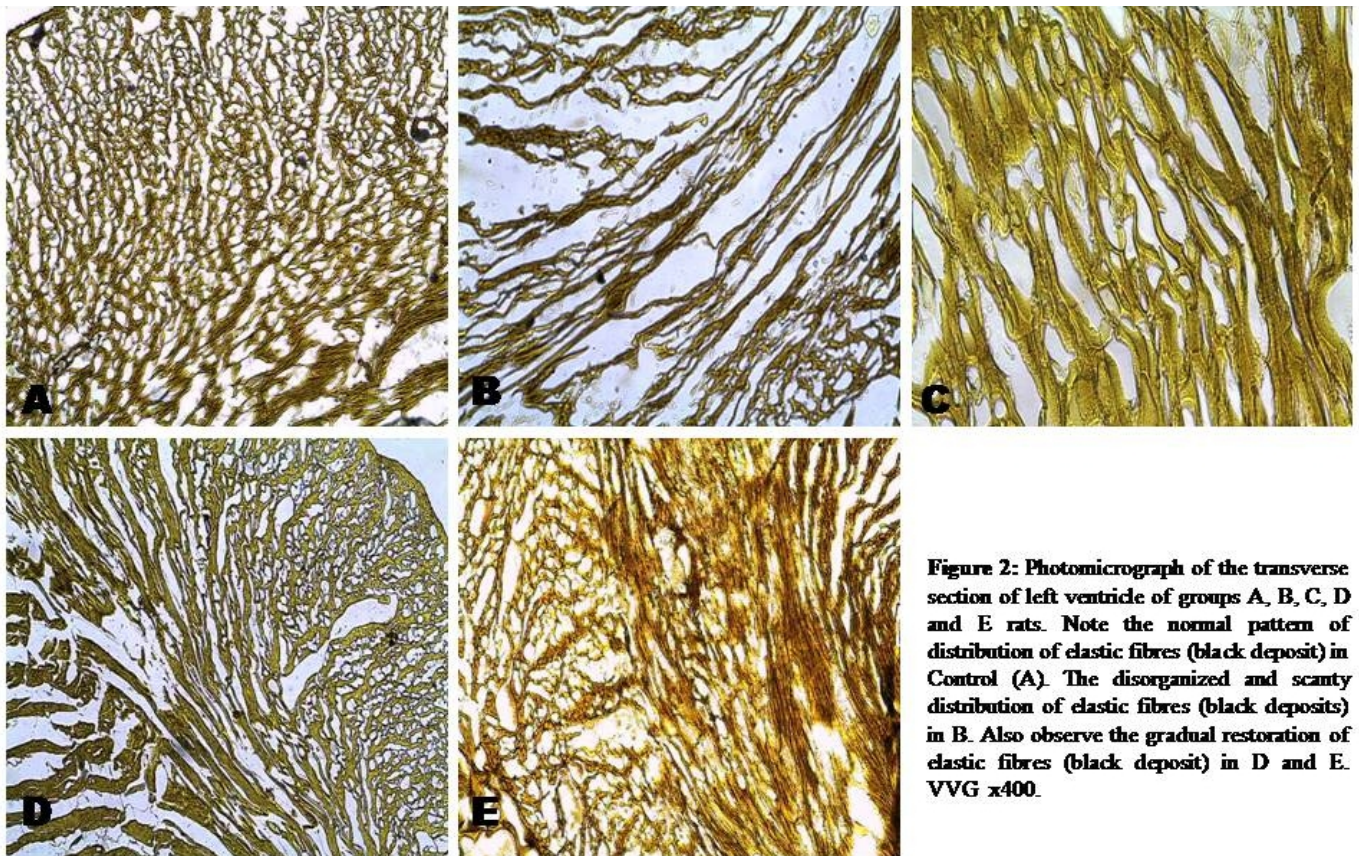
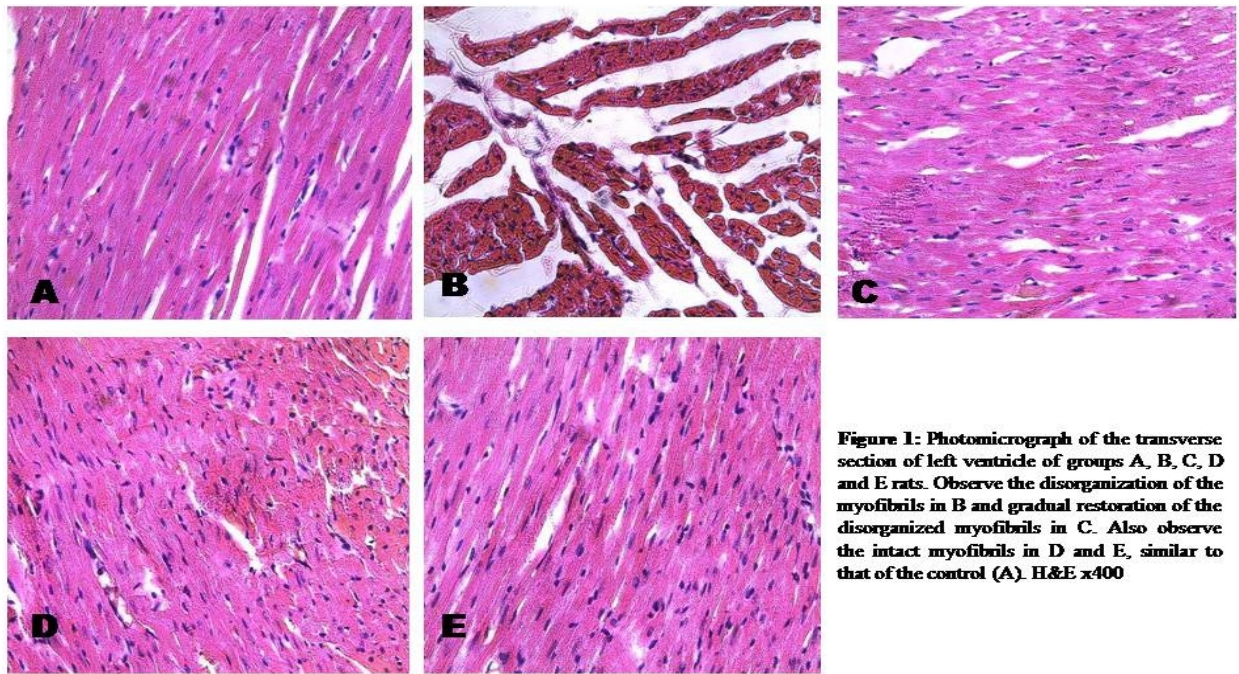
At the end of the experimental period, all the animals were physically observed and anesthetized by chloroform inhalation. A midline incision was performed at the thoracic region. The heart was harvested and the left ventricle was excised, fixed in 10 % Formol saline and 10% neutral buffered formalin for 72 hours.

Histology, Histochemistry and Photomicrography:

Tissues were processed routinely in paraffin wax embedding. Sections of 6µm thick were cut and stained using hematoxylin and eosin (H&E) procedure, Verhoeff - Van Gieson (VVG) staining procedure for elastic fibre and Periodic Acid Schiff (PAS) staining procedure for demonstration of glycogen. The sections were examined under a LEICA research microscope (LEICA Dm750, Switzerland) with a digital camera attached (LEICA ICC50). Digital photomicrographs of stained sections were taken.

RESULTS

The control section showed a normal histological outline of the left ventricle characterized by well organized myofibrils as against disorganized myofibrils in the diabetic group (Fig. 1). These alterations were greatly attenuated with administration of *M. charantia* and glimepiride for four weeks. This effect was better improved with *M. charantia* than glimepiride. Withdrawal of extract only showed mild improvement in the histological outline. Evidences from Verhoeff-vanGieson stain revealed that the elastic fibers were greatly reduced in the myocardium of the left ventricle in the diabetic group. With the commencement of extract administration and glimepiride for four weeks, there were significant improvements in the restoration of elastic fibers as evident in the staining intensity. Withdrawal of extract treatment slightly improves the deposition of elastic fibers (Fig. 2). Histochemical findings showed accumulation of glycogen deposit in the left ventricle of diabetic rats (Fig. 3). Treatment of diabetic rats with *M. charantia* and glimepiride for four weeks reduced the PAS positive staining intensity. The group treated with *M. charantia* for two weeks, revealed a mild reduction in the PAS positive staining intensity.



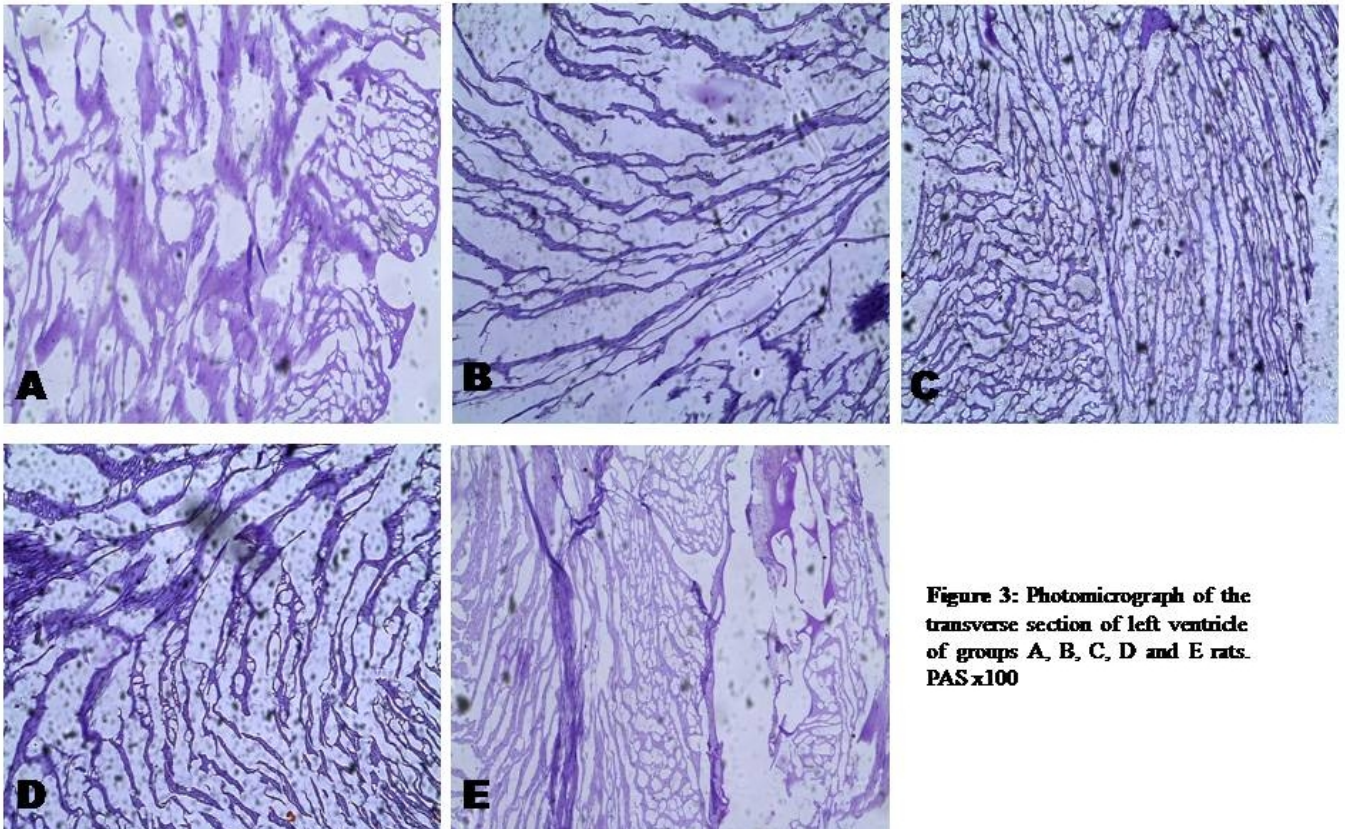


Figure 3: Photomicrograph of the transverse section of left ventricle of groups A, B, C, D and E rats. PAS x100

DISCUSSION

Diabetes has been reported to be associated with profound alterations in biochemical and normal histology leading to an increased risk of coronary heart disease^{22,23,24,25}. Diabetic cardiomyopathy characterized by diastolic dysfunction and left ventricular hypertrophy is usually the terminal condition of heart in diabetes²⁶.

Histologically, there was evidence of architectural alteration in the myocardium of diabetic animals. This observation is consistent with previous study²⁷. These effects were abrogated with the administration of *M. charantia* extract and glimepiride for four weeks. This observation may have been possible due to a reduction in blood glucose level leading to enhanced peripheral glucose utilization. Tripathi and Chandra et al²⁸ also reported that *M. charantia* extracts potentiate the insulin effect by rejuvenation of damaged pancreatic β cell.

Distributions of elastic fibres were observed to be sparsely distributed as evident by the staining intensity in the left ventricle of diabetic rats when compared with the control group. This observation suggests a reduction in the tensile strength and elasticity of the heart. Administration of *M. charantia* and glimepiride gradually restored the integrity of these fibres there by reducing the susceptibility to cardiovascular complications such as stroke syndrome, myocardial infarction and hypertension²⁶. Histochemical findings suggest accumulation of glycogen deposit in diabetic rats. In general, glycogen accumulation is an anatomic indicator of a deranged carbohydrate metabolism in alloxan-diabetes which probably reflects the hyperglycaemia²⁹. Glycogen is synthesized in the body in a reaction beginning with glucose-1- phosphate which is converted to uridine diphosphate glucose (UDPG) and then to glycogen in reactions catalysed by the enzyme UDPG glycogen-transferase, and the branching enzyme (amylo-1, 4 $\rightarrow$ 1, 6 transglucosidase). The reduction in the PAS staining intensity is reflective of reduced glycogen content in the group treated with *M. charantia* and glimepiride for four weeks²⁹. All these evidences suggest cardio-protective effects of *M. charantia* against anatomical derangements observed in the diabetic group. Thus the present results showed that the methanolic extract of *M. charantia* has a promising ameliorative effects on the associated cardiac complications implicated in STZ induced diabetes in rats.

REFERENCES

- Vats V, Yadav SP, Grover JK. Ethanolic extract of *Ocimum sanctum* leaves partially attenuates streptozotocin-induced alterations in glycogen content and carbohydrate metabolism in rats. *J. Ethnopharmacol* 2004; 90: 155-160.
- Jarrett RJ. Cardiovascular disease and hypertension in diabetes mellitus. *Diabetes/metabolism reviews* 1989; 5: 547-558.
- Grundy SM, Benjamin IJ, Burke GL, Chait A, Eckel RH, Howard BV et al. *Diabetes and cardiovascular disease: a statement for healthcare professionals from the American Heart Association. Circulation* 1999; 100: 1134-1146.
- The Diabetes Control and Complications Trial Research Group. (TDCCTRG) The effect of intensive treatment of diabetes on the development and progression of long-term complications in insulin-dependent diabetes mellitus. *N Engl J Med* 1993; 329(14): 977-986.
- Malone JJ, Schocken DD, Morrison AD, Gilbert-Barness E. Diabetic cardiomyopathy and carnitine deficiency. *Jour Diab Comp* 1999; 13: 86-89.
- Hoit BD, Castro C, Bultron G, Knight S, Matlib MA. "Noninvasive evaluation of cardiac dysfunction by echocardiography in streptozotocin-induced diabetic rats". *J. Card Fail* 1999;5: 324-333.
- Van Buren T, Schiereck P, De Ruiter GJW, Gispen WH, De Wildt DJ. Vagal efferent control of electrical properties of the heart in experimental diabetes. *Acta Diabetologica* 1998; 35: 19-25.
- Aring AM, Jones DE, Falko JM. Evaluation and prevention of diabetic Neuropathy. *AM FAM Physician* 2005; 71:2123- 2130.
- Malone JJ, Lowitt S, Korthals JK, Salem A, Miranda C. The effect of hyperglycemia on nerve conduction and structure is age dependent. *Diabetes* 1996; 45: 209-215.
- Mamoulakis D, Galanakis E, Dionyssopoulou E, Evangeliou A, Sbyrakis S. Carnitine deficiency in children and adolescents with type 1 diabetes. *J. Diabetes. Complications* 2004; 18: 271-4.
- Braca A, Siciliano T, D'Arrigo M, Germanò MP. Chemical composition and antimicrobial activity of *Momordica charantia* seed essential oil. *Fitoterapia* 2008; 79: 123-125.
- Platel K, Srinivasan K. Plant foods in the management of diabetes mellitus: vegetables as potential hypoglycaemic agents. *Nahrung* 1997; 41(2); 68-74.
- Raman A, Lau C. Anti-diabetic properties and phytochemistry of *Momordica charantia* L. (Cucurbitaceae). *Phytomedicine* 1996; 2: 349-662.
- Nicholas LB, Kolb Y, Prinssen EP. A combined marble burying locomotor activity test in mice: a practical screening test with sensitivity to different classes of anxiolytics and antidepressants. *Eur J Pharmacol* 2006; 547: 106 -115.
- Akhtar MS, Athar MA, Yaqub M. Effect of *Momordica charantia* on blood glucose level of normal and alloxan-diabetic rabbits. *Planta Med* 1981; 42: 205-212.
- Sarkar S, Pranava M, Marita R. Demonstration of the hypoglycaemic action of *Momordica charantia* in a validate animal model of diabetes. *Pharmacol. Res* 1996; 33: 1-4.
- Miura T, Ichiki H, Hashimoto I, Iwamoto N, Kato M, Kubo M. et al. Impairment of insulin-stimulated GLUT4 translocation in skeletal muscle and adipose tissue in the Tsumura Suzuki obese diabetic mouse: a new genetic animal model of type 2 diabetes. *Eur J Endocrinol* 2001; 45: 785-790.
- Nagasawa H, Watanabe K, Inatomi H. Effects of bitter melon (*Momordica charantia*) or ginger rhizome (*Zingiber officinale* Rosc.) on spontaneous mammary tumorigenesis in SHN mice. *Am. J. Clin. Me* 2002; 30(2-3); 195-205.
- Tan FJ, Fire AZ, Hill RB. Regulation of apoptosis by C. elegans CED-9 in the absence of the C-terminal transmembrane domain. *Cell Death Differ* 2007; 14: 1925-1935.
- National institute of health guide for the care and use of laboratory animals DHEW publication (NIH) revised office of science and health report DRR/NIH Bethesda; USA, 1996, 1-10.
- Mir SH, Darzi MM, Ahmad F, Chishti MZ, Mir MS. The Influence of Glimepiride on the Biochemical and Histomorphological Features of Streptozotocin-Induced Diabetic Rabbits. *Pakistan Journal of Nutrition* 2008; 7(3); 404-407.
- Huang TM, Lee-Huang S, Huang PL, Chen HC. Studies on antiviral activity of the extract of *Momordica charantia* and its active principle." *Virologica* 1988; 5(4); 367-73.
- Fontbonne A, Eschwege E, Cambien F, Richard JL, Ducimetiere P, Thibault N et al. Hypertriglyceridaemia as a risk factor of coronary heart disease mortality in subjects with impaired glucose tolerance or diabetes. *Diabetologia* 1989; 32: 300-304.
- Motta M, Giugno I, Bosco S, Pistone G, Ruello P, Maugeri D. et al. Serum lipoprotein(a) changes in acute myocardial infarction. *Panminerva Medica* 2001; 43: 77-80.
- Maghrani M, Lemhadri A, Zeggwagh NA, El Amraoui M, Haloui M, Jouad H et al. Effects of an aqueous extract of *Triticum repens* on lipid metabolism in normal and recent-onset diabetic rats. *Journal of Ethnopharmacology* 2004; 90: 331-337.
- Raev DC. Which left ventricular function is impaired earlier in the evolution of diabetic cardiomyopathy? An echocardiographic study of young type I diabetic patients. *Diabetes Care* 1994, 17(7):633-639.
- Komolafe OA. Effects of streptozotocin induced diabetes mellitus on the aorta, pulmonary trunk and left ventricle of *rattus norvegicus* M.Sc thesis, Obafemi Awolowo University, Ile-ife, Osun State, Nigeria, 2009.
- Tripathi UN, Chandra D. The plant extracts of *Momordica charantia* and *Trigonella foenum graecum* have antioxidant and anti-hyperglycemic properties for cardiac tissue during diabetes mellitus. *Oxidative Medicine and Cellular Longevity* 2009, 2(5): 290-296
- Newell FW and Kurimoto S. Histochemistry of the retina in the alloxan-diabetic rat. *Br J Ophthalmol* 1963, 47; 596-600.