

Journal of Pharmaceutical and Scientific Innovation

www.jpsionline.com

Research Article

SYNTHESIS OF N-METHYLATED ANALOG OF AMIDOMYCIN: A POTENT ANTIMICROBIAL CYCLOHEPTAPEPTIDE

Himaja M. 1*, Ogale Sachin Ramachandra Rao², Moonjit Das 1 and Asif Karigar 3

¹Pharmaceutical Chemistry Division, School of Advanced Sciences, VIT University, Vellore, Tamil Nadu, India

²Department of Pharmaceutical Chemistry, Nitte Gulabi Shetty Memorial Institute of Pharmaceutical Sciences, Nanthoor, Mangalore, India

³Department of Pharmaceutical Analysis, Maratha Mandal's College of Pharmacy, Belgaum, Karnataka, India

*Corresponding Author Email: drmhimaja@gmail.com

DOI: 10.7897/2277-4572.033142

 $Published\ by\ Moksha\ Publishing\ House.\ Website\ www.mokshaph.com$

All rights reserved.

Received on: 05/04/14 Revised on: 02/05/14 Accepted on: 05/06/14

ABSTRACT

N-methylated analog of Amidomycin, was synthesized by solution phase peptide synthesis using dicyclohexylcarbodiimide (DCC) as the coupling agent and triethylamine (TEA) as the base. The structure of the compound was confirmed by IR, ¹H NMR, ¹³C NMR, FABMASS and elemental analysis. The synthesized cyclic peptide was evaluated for Minimum Inhibitory Concentration (MIC) against four strains of bacteria and three strains of fungi. It was found to be active against both bacteria and fungi from 50-100 µg.

Keywords: Amidomycin, solution phase peptide synthesis, DCC, MIC.

INTRODUCTION

Amidomycin, a cyclic depsipeptide, was produced by a Streptomyces species. In the year 1957, Vining L.C. et al. 1 first isolated it and elucidated its structure. Synthesis of amidomycin was carried out by Shemyakin M. M. et al. in the year 1963². It is an antibiotic which is primarily active against yeasts. However, no further studies were carried out on Amidomycin because of the complexity of the synthesis of the alternate amide and ester linkages and usage of Damino acid which is very costly. Hence simple analogues were designed with only amide linkages, thus making synthesis easier and cost effective by incorporating L-amino acids. N-methylated amino acids are commonly found in naturally occurring peptide antibiotics. The methylation of Natom eliminates the hydrogen, responsible for cleavage of peptide bonds. The hydrogen bonding pattern of peptide containing these amino acids is different from that of unmethylated forms.³⁻⁵ N-methylated cyclic peptides is found to possess cytotoxic and antimicrobial activity. A review of the structures of cyclic peptides exhibiting antimicrobial activity showed presence of at least one D-amino acid and/or N-methylated amino acid units in the molecule. Hence the cyclic octapeptide analogue of amidomycin have been designed containing one N-methyl-L-valine and rest L-valine units. In order to carry out the synthesis, the cyclic octapeptide was disconnected into four dipeptide units. The dipeptides were prepared from the respective protected amino acids. The amino group was protected with tertiary Butyloxycarbonyl (Boc-) group and the carboxyl group was protected by converting it into the methyl ester. The Bocamino acids were coupled with the amino acid methyl ester hydrochlorides by dicyclohexylcarbodiimide (DCC) as the coupling agent and triethylamine (TEA) as the base to get the protected dipeptides. The dipeptides were appropriately de protected and coupled to get the octapeptides, which were finally cyclised by p-nitrophenyl ester method using highdilution technique to get the cyclic octapeptide.

MATERIALS AND METHODS

All the reactions requiring anhydrous conditions were conducted in dried apparatus. All the reactions were magnetically stirred unless otherwise stated. Organic extracts were dried over anhydrous sodium sulphate. Melting points were determined by capillary method and were uncorrected. Amino acids, di-tert-butylpyrocarbonate, trifluoroacetic acid and triethylamine were obtained from Spectrochem Ltd. Mumbai, India. DCC, Diethyl ether, Methanol and Chloroform was obtained from AVRA. IR spectra were recorded on Jasco FT/IR-5300 IR spectrometer using a thin film supported on KBr pellets for solids and chloroform as a solvent for semisolids. The values are reported as v_{max} (cm⁻¹). ¹H NMR spectra were recorded on Bruker JOEL (400 MHz) NMR spectrometer. The spectra were obtained in CDCl₃ and the chemical shift values are reported as values in ppm relative to TMS ($\delta = 0$) as internal standard. FABMASS spectra were recorded on a Joel Sx 102/DA-6000 mass spectrometer using xenon as the carrier gas. The spectra were recorded at room temperature; m-nitrobenzyl alcohol was used as the matrix. The protection of amino and carboxyl group and their de protection were done by standard procedures.6-9

N-methylation of Boc-amino acid

Boc-amino acid (0.663 g, 2.0 mmol) was dissolved in dry THF (20 ml) and cooled to 0°C. To this NaH (2.88 g, 12 mmol) and MeI (0.852 g, 6.0 mmol) were added and stirred overnight at room temperature. The reaction mixture was diluted with ether (20 ml), washed with sat. NH₄Cl (10 ml), 20 % Na₂S₂O₃ solution and sat. NaCl solution (10 ml); Organic layer was dried and concentrated. Using the above method following N-methyl amino acid was prepared.

Preparation of Dipeptides

Amino acid methyl ester hydrochloride (10 mmol) was dissolved in chloroform (CHCl₃) (20 ml). To this, TEA (4 ml, 28.7 mmol) was added at 0°C and the reaction mixture was stirred for 15 minutes. Boc-amino acid (10 mmol) in CHCl₃

(20 ml) and DCC (10 mmol) were added with stirring. After 36 h, the reaction mixture was filtered and the residue was washed with CHCI $_3$ (30 ml) and added to the filtrate. The filtrate was washed with 5 % NaHCO $_3$ (20 ml), 5 % HCl (20 ml) and distilled H $_2$ O (20 ml). The organic layer was dried over anhydrous Na $_2$ SO $_4$, filtered and evaporated in a vacuum. The residue was purified by re crystallization from CHCl $_3$. Boc-L-Val-Val-OMe and Boc-N-methyl-L-Val-L-Val-OMe was prepared in this manner.

Preparation of Tetrapeptides

The de protected dipeptide units were coupled using DCC/TEA to get the protected tetrapeptide by the procedure similar to that of the dipeptides. Boc-L-[Val-Val-Val-Val]-OMe and Boc-N-methyl-L-Val-L-[Val-Val-Val]-OMe were synthesized in this manner.

Preparation of linear octapeptide

The Boc-group of the tetrapeptides Boc-L-[Val-Val-Val-Val]-OMe was removed and the ester group of the

tetrapeptide Boc-L-Val-L-Val-N-methyl-L-Val-L-Val-OMe was de protected. Both the de protected units were coupled to get the linear octapeptide.

Preparation of Cyclic octapeptide

The cyclisation of the linear octapeptide unit was carried out by the p-nitrophenyl ester⁶ with certain modifications. The ester group of the linear fragment was removed and the pnitrophenyl ester group was introduced by stirring it for 12 h in CHCl₃ with p-nitrophenol at 0°C. The reaction mixture was washed several times with saturated NaHCO3 until the unreacted p-nitrophenol was removed completely and washed with 5 % HCl to get Boc-peptide-pnp ester. The Boc-group was also removed, CHCl3 and pyridine was added and the reaction mixture was kept at 0°C for 10 days. The mixture was finally washed with 5 % HCl, dried and evaporated in vacuum to get the cyclised product, which was then re crystallized from CHCl₃/n-hexane (Scheme

Cyclo-N-methyl-L-Val-L-[Val-Val-Val-Val-Val-Val-Val]

Scheme 1

Determination of Minimum Inhibitory Concentration (MIC)

The MIC of the cyclic peptide was determined by the serial tube dilution technique 10-12 against two strains of Gram positive bacteria (Staphylococcus aureus and Bacillus subtilis), two strains of Gram negative bacteria (Escherichia coli and Pseudomonas aeruginosa) and three strains of fungi (Candida albicans, Asperigillus flavus and Asperigillus fumigatus). 4 mg of the sample was dissolved in 2 ml of sterile dimethyl formamide (DMF) to obtain stock solution having concentration of 200 µg/ml. In serial dilution technique, 1 ml prepared stock solution was transferred to test tube containing 1 ml nutrient broth medium for bacterial cultures and 1 ml Potato Dextrose Broth (PDB) for fungal

cultures to give concentration 100 µg/ml from which 1 ml was transferred to another test tube containing 1 ml of broth medium to give concentration 50 µg/ml and so on up to concentration 6.25 µg/ml. After preparation of suspension of test organisms (10 organisms per ml), 1 drop of suspension (0.02 ml) was added to each broth dilution; a positive control was prepared in a similar way except that the test compound was not added. A negative control was prepared without the test compound and the test organisms. Tubes inoculated with bacterial cultures were incubated aerobically at 37°C for 24 hours and tubes inoculated with fungal cultures were incubated aerobically at 25°C for 48 hours. The tubes were presence/absence of observed for the growth.

Concentration of the solutions of the Test Compound

Concentration of the stock solution = $4 \text{ mg/2 ml} (200 \text{ µg/ml})$											
Tube number	I	II	III	IV	V	VI					
Concentration µg/ml	6.25	12.5	25	50	100	200					

RESULTS AND DISCUSSION

Physical Data and Spectral Analysis

Cyclo-N-methyl-L-Val-L-[Val-Val-Val-Val-Val-Val-Val-Val]

Yield 81.35 %; light brown semisolid; IR spectrum (v/cm⁻¹): 3287.4 cm⁻¹ (br. s, -NH Stretch), 2932 cm⁻¹ (s, -CH stretch), 1647.3 cm⁻¹ (s, C=O stretch), 1557 cm⁻¹ (s, -NH bend), 1452.8 cm⁻¹ (s, NH bend); ¹H NMR spectrum (δ, ppm): 8.05(1H, br. s, -NH), 7.4(3H, br. s, -NH), 6.9(1H, br.s, -NH), 6.45(3H, br. s, -NH), 4.6(2H, m, α-H), 4.4(1H, m, α-H), 4.25(2H, m, α-H), 4.1(2H, m, α-H), 4.0(1H, m, α-H), 2.2(3H, s, -NCH₃), 1.4-1.0(8H, m, β-H), 1.0-0.9(48H, m, -CH₃); ¹³C NMR: (75.467MHz, CDCl₃): 170.4(C=O of Val), 59.9(α-C), 52.7(α-C), 52.0 (α-C), 48.7(α-C), 33.8 (β-C),

33.5 (β-C), 32.8 (β-C), 32.2 (β-C), 31.0 (β-C), 30.5 (β-C), 30.2 (β-C), 29.6 (β-C), 25.9 (CH₃), 25.6 (CH₃), 25.4 (CH₃), 25.2 (CH₃), 24.9 (CH₃), 24.7 (CH₃), 19.2 (CH₃), 18.8 (CH₃), 17.8 (CH₃); FABMASS: m/z (M + 1)⁺ = 809; Elemental Analysis: M. F. = C₄₁H₇₄N₈O₈, M. W. = 809, Found (Cal) % C: 64.43 (65.13), % N: 9.97 (10.0).

Minimum Inhibitory Concentration (MIC)

The synthesized cyclic peptides were evaluated for antibacterial and antifungal activities (MIC) from 200 μg to 6.25 μg . The cyclic peptide showed activity from 50 μg to 200 μg against all bacterial and fungal strains. The results of the MIC are given in Table 1 and Table 2.

Table 1: Minimum Inhibitory Concentration for Antibacterial Activity

Comp. No↓		Presence/absence of growth																						
Organism →	S. aureus						B. subtilis					E. coli						P. aeruginosa						
Dilution→	I	II	II I	I V	V	V I	I	II	II I	I V	V	V I	I	II	II I	I V	V	V I	Ι	II	II I	I V	V	V I
CP-2	_	-	_	+	+	+	_	_	_	+	+	+	_	_	_	+	+	+	_	_	_	+	+	+

^{&#}x27;+' indicates presence of growth, '-' indicates absence of growth

Table 2: Minimum inhibitory concentration for antifungal activity

Compd. No.↓		Presence/absence of growth																		
Organism →			C. all	bicans			A. flavus							A. fumigatus						
Dilution →	I	II	III	IV	V	VI	I	II	III	IV	V	VI	I	II	III	IV	V	VI		
CP-2	-	ı	-	+	+	+	-	ı	+	+	+	+	-	-	-	+	+	+		

^{&#}x27;+' indicates presence of growth, '-' indicates absence of growth

CONCLUSION

The N-methylated cyclic octapeptide could be conveniently and efficiently synthesized by the prescribed scheme with good yields. The structure of the cyclic peptide was confirmed by IR, ¹H NMR, ¹³C NMR, FABMASS and elemental analysis. The compounds were screened for the Minimum Inhibitory Concentration (MIC) against four strains of bacteria and three strains of fungi and it was found to be active against both bacteria and fungi from 50-200 μg.

ACKNOWLEDGEMENTS

We are grateful to the DST/VIT-SIF Lab, School of Advanced Sciences, VIT University, Vellore, India for the support of this research.

REFERENCES

- Vining LC, Taber WA. Amidomycin, a new antibiotic from a Streptomyces species Can J Chem 1957; 35: 1109. http://dx.doi. org/10.1139/v57-151
- Shemyakin MM, Vinogradova EI, Feigina MY, Aldanova NA. On the structure of amidomycin and valinomycin, Tetrahedron Letters 1963; 4(6): 351-356. http://dx.doi.org/10.1016/S0040-4039(01)90635-5

- Belagali SL, Matthew T, Himaja M, Kocienski PA. A highly efficient method of N-methylation for the amino acid derivatives. Indian J Org Chem 1995; 34: 44.
- Himaja M, Vani G, Ramana MV, Karigar A, Munirajasekhar D. Synthesis of Glabrin A and its N-methylatedanalog as potent insecticidal agents. Int. J. Res. Ayu. Pharm 2012; 3(1): 91-94.
- Das P, Himaja M. Design and synthesis of 4-[2'-(5'- nitro)] imidazolyl benzoyl (n-methyl) amino acids and peptides. Int J Drug Dev Res 2010; 2(2): 364-370.
- Dahiya R, Gautam H. Total synthesis and antimicrobial activity of a natural cycloheptapaptide of marine origin. Marine Drugs 2010; 8: 2384. http://dx.doi.org/10.3390/md8082384
- Bodanszky M, Bodanszky A. Practice of Peptide Synthesis. Springer-Verlag, New York; 1984. p. 78. http://dx.doi.org/10.1007/978-3-642-96835-8
- Das M, Agarwal DS, Sharma SK, Das P, Rout PK. Synthesis, docking studies and free radical scavenging activity of the linear tetrapeptide VFPF. Int. Res. J. Pharm 2012; 3(6): 138.
- Das M, Himaja M. Synthesis, antioxidant and anthelmintic activity of the linear tetrapeptide L-(Leu-Pro-Gly)-D-Ala (LPGA). Int. J. Pharm. Pharm. Sci 2013; 5 (Suppl 3): 713.
- Ekramul IM, Ekramul HM, Ashiik MM. Cytotoxicity and antibacterial activity of *Sida rhombifolia* (Malvaceae) grown in Bangladesh. Phyt Res 2003; 17(8): 978-975.
- 11. Marjorie CM. Plant products as antimicrobial agents. Clin Microbiol Rev 1999; 12(4): 564-582.
- Roy S, Choudhury MD, Paul SB. In vitro Antibacterial activity of Alocasia decipiens schott. Int J Pharm Pharm Sci 2013; 5 Suppl 1: 155-157

Source of support: Nil, Conflict of interest: None Declared

QUICK RESPONSE CODE

ISSN (Online): 2277 -4572

Website http://www.jpsionline.com

How to cite this article:

Himaja M., Ogale Sachin Ramachandra Rao, Moonjit Das and Asif Karigar. Synthesis of N-methylated analog of amidomycin: A potent antimicrobial cycloheptapeptide. J Pharm Sci Innov. 2014;3(3):217-220 http://dx.doi.org/10.7897/2277-4572.033142